# Package 'MarZIC'

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Type Package

<b>Title</b> Marginal Mediation Effects with Zero-Inflated Compositional Mediator	
Version 1.0.1	
<b>Description</b> A way to estimate and test marginal mediation effects for zero-inflated compositional mediators. Estimates of Natural Indirect Effect (NIE), Natural Direct Effect (NDE) of each taxon, as well as their standard errors and confident intervals, were provided as outputs. Zeros will not be imputed during analysis. See Wu et al. (2022) <doi:10.3390 genes13061049="">.</doi:10.3390>	
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MarZIC

Marginal Mediation Model for Zero-Inflated Compositional Mediators

#### Description

MarZIC is used for calculating marginal mediation effects for zero-inflated compositional mediators. For microbiome data, the marginal outcome model for the *j*th taxon (or OTU, ASV) is:

$$Y = \beta_0 + \beta_1 M_j + \beta_2 1_{M_i > 0} + \beta_3 X + \beta_4 X 1_{M_i > 0} + \beta_5 X M_j + \epsilon$$

where  $1_{()}$  is indicator function, X is the covariate of interest and  $M_j$  is the relative abundance of the jth taxon. The probability of  $M_j$  being structure zero (ie, true zeros) is:

$$\log(\frac{\Delta_j}{1 - \Delta_j}) = \gamma_0 + \gamma_1 X$$

The mean of  $M_j$  in compositional structure is modeled as:

$$\log(\frac{\mu_j}{1-\mu_j}) = \alpha_0 + \alpha_1 X$$

Typically, users just need to feed the first seven inputs to the function: 'MicrobData', 'CovData', 'lib\_name', 'y\_name', 'x\_name', 'conf\_name' and 'taxa\_of\_interest'.

### Usage

```
MarZIC(
  MicrobData,
  CovData,
  lib_name
  y_name,
  x_name,
  conf_name = NULL,
  x4_{inter} = TRUE,
  x5_inter = TRUE,
  taxa_of_interest = "all",
  mediator_mix_range = 1,
  transfer_to_RA = TRUE,
  num_cores = max(detectCores() - 2, 1),
  adjust_method = "fdr",
  fdr_rate = 0.05,
  taxDropThresh = 0.8,
  taxDropCount = 4 * (length(conf_name) + 2),
  zero_prop_NIE2 = 0.1,
  zero_count_NIE2 = 4 * (length(conf_name) + 2),
  SDThresh = 0.05,
  SDx = 0.05,
  SDy = 0.05
)
```

#### **Arguments**

MicrobData A dataset contains microbiome data. The microbiome data could be relative

abundance or absolute abundance. Subjects with missing value will be removed

during analysis.

CovData A dataset contains outcome, library size and covariates.

lib\_nameName of library size variable within colData.y\_nameName of outcome variable within colData.x\_nameName of covariate of interest within colData.

conf\_name Name of confounders within colData. Defaule is NULL, meaning no con-

founder.

x4\_inter Whether to include the interaction term  $\beta_4$ . Default is TRUE. x5\_inter Whether to include the interaction term  $\beta_5$ . Default is TRUE.

taxa\_of\_interest

A character vector for taxa names indicating taxa that should be analyzed. Default is "all", meaning all taxa should be included into analysis.

mediator\_mix\_range

Number of mixtures in mediator. Default is 1, meaning no mixture.

transfer\_to\_RA Logical variable indicating whether the microbiome data should be transferred

to relative abundance. Default is TRUE. If TRUE, microbiome data will be

rescaled by its row sum.

num\_cores Number of CPU cores to be used in parallelization task.

adjust\_method P value adjustment method. Same as p.adjust. Default is "fdr".

fdr\_rate FDR cutoff for significance. Default is 0.05.

taxDropThresh The threshold of dropping taxon due to high zero percentage. Default is 0.9,

meaning taxon will be dropped for analysis if zero percentage is higher than

90%.

taxDropCount The threshold of dropping taxon due to not enough non-zero observation counts.

Default is 4 \* (length(conf\_name)+2), meaning taxon will be dropped if non-

zero observation is less than four times of number of covariates plus 1.

zero\_prop\_NIE2 The threshold of zero percentage for calculating NIE2. Default is 0.1, meaning

NIE2 will be calculated for taxon with zero percentage greater than 10%.

zero\_count\_NIE2

The threshold of zero counts for calculating NIE2. Default is 4 \* (length(conf\_name)+2),

meaning NIE2 will be calculated for taxon with zero counts greater than four

times of number of covariates plus 1.

SDThresh The threshold of dropping taxon due to low coefficient of variation (CV) to avoid

constant taxon. Default is 0.05, meaning any taxon has CV less than 0.05 will

be dropped.

SDx The threshold of stopping analysis due to low CV of covariate of interest. De-

fault is 0.05, meaning when CV of covariate of interest is less than 0.05, the

analysis will be stopped.

SDy The threshold of stopping analysis due to low CV of outcome. Default is 0.05,

meaning when CV of outcome. is less than 0.05, the analysis will be stopped.

#### Value

A 'list' of '4' datasets containing the results for 'NIE1', 'NIE2', 'NDE', and 'NIE'. Each dataset has row representing each taxon, 6 columns for 'Estimates', 'Standard Error', 'Lower bound for 95 'Adjusted p value', 'Significance indicator'.

#### References

Wu et al.(2022) MarZIC: A Marginal Mediation Model for Zero-Inflated Compositional Mediators with Applications to Microbiome Data. Genes 2022, 13, 1049.

#### **Examples**

```
library(MarZIC)
## A make up example with 2 taxon and 20 subjects.
set.seed(1)
nSub <- 20
nTaxa <- 2
## generate covariate of interest X
X \leftarrow rbinom(nSub, 1, 0.5)
## generate confounders
conf1<-rnorm(nSub)</pre>
conf2<-rbinom(nSub,1,0.5)</pre>
## generate mean of each taxon. All taxon are having the same mean for simplicity.
mu \leftarrow exp(-5 + X + 0.1 * conf1 + 0.1 * conf2) /
 (1 + \exp(-5 + X + 0.1 * conf1 + 0.1 * conf2))
phi <- 10
## generate true RA
M_taxon<-t(sapply(mu,function(x) dirmult::rdirichlet(n=1,rep(x*phi,nTaxa))))</pre>
P_{zero} \leftarrow exp(-3 + 0.3 * X + 0.1 * conf1 + 0.1 * conf2) /
 (1 + \exp(-3 + 0.3 * X + 0.1 * conf1 + 0.1 * conf2))
non_zero_ind <- t(sapply(P_zero,function(x) 1-rbinom(nTaxa,1,rep(x,nTaxa))))</pre>
True_RA<-t(apply(M_taxon*non_zero_ind,1,function(x) x/sum(x)))</pre>
## generate outcome Y based on true RA
Y <-1 + 100 * True_RA[,1] + 5 * (True_RA[,1] > 0) + X + conf1 + conf2 + rnorm(nSub)
## library size was set to 10,000 for all subjects for simplicity.
libsize <- 10000
## generate observed RA
observed_AA <- floor(M_taxon*libsize*non_zero_ind)</pre>
observed_RA <- t(apply(observed_AA,1,function(x) x/sum(x)))</pre>
colnames(observed_RA)<-paste0("rawCount", seq_len(nTaxa))</pre>
CovData <- cbind(Y = Y, X = X, libsize = libsize, conf1 = conf1, conf2 = conf2)
```

```
## run the analysis
res <- MarZIC(
   MicrobData = observed_RA,
   CovData = CovData,
   lib_name = "libsize",
   y_name = "Y",
   x_name = "X",
   conf_name = c("conf1","conf2"),
   taxa_of_interest = NULL,
   num_cores = 1,
   mediator_mix_range = 1
)
}</pre>
```

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